

---

*This copy is for your personal, non-commercial use only.*

---

**If you wish to distribute this article to others**, you can order high-quality copies for your colleagues, clients, or customers by [clicking here](#).

**Permission to republish or repurpose articles or portions of articles** can be obtained by following the guidelines [here](#).

**The following resources related to this article are available online at [www.sciencemag.org](http://www.sciencemag.org) (this information is current as of April 29, 2011 ):**

**Updated information and services**, including high-resolution figures, can be found in the online version of this article at:

<http://www.sciencemag.org/content/331/6018/768.full.html>

**Supporting Online Material** can be found at:

<http://www.sciencemag.org/content/suppl/2011/02/08/331.6018.768.DC1.html>

A list of selected additional articles on the Science Web sites **related to this article** can be found at:

<http://www.sciencemag.org/content/331/6018/768.full.html#related>

This article **cites 27 articles**, 18 of which can be accessed free:

<http://www.sciencemag.org/content/331/6018/768.full.html#ref-list-1>

This article has been **cited by** 1 articles hosted by HighWire Press; see:

<http://www.sciencemag.org/content/331/6018/768.full.html#related-urls>

This article appears in the following **subject collections**:

Medicine, Diseases

<http://www.sciencemag.org/cgi/collection/medicine>

## References and Notes

1. A. Ciechanover, *Nat. Rev. Mol. Cell Biol.* **6**, 79 (2005).
2. A. Hershko, A. Ciechanover, *Annu. Rev. Biochem.* **67**, 425 (1998).
3. A. L. Schwartz, A. Ciechanover, *Annu. Rev. Med.* **50**, 57 (1999).
4. K. I. Nakayama, K. Nakayama, *Nat. Rev. Cancer* **6**, 369 (2006).
5. J. M. Pratt et al., *Mol. Cell. Proteomics* **1**, 579 (2002).
6. U. Alon, *An Introduction to Systems Biology* (Chapman & Hall, New York, 2007).
7. R. T. Schimke, D. Doyle, *Annu. Rev. Biochem.* **39**, 929 (1970).
8. H. C. Yen, Q. Xu, D. M. Chou, Z. Zhao, S. J. Elledge, *Science* **322**, 918 (2008).
9. J. Monod, A. M. Pappenheimer Jr., G. Cohen-Bazire, *Biochim. Biophys. Acta* **9**, 648 (1952).
10. Materials and methods are available as supporting material on Science Online.
11. A. A. Cohen et al., *Science* **322**, 1511 (2008).
12. A. Sigal et al., *Nat. Protoc.* **2**, 1515 (2007).
13. T. Surrey et al., *Proc. Natl. Acad. Sci. U.S.A.* **95**, 4293 (1998).
14. O. Tour, R. M. Meijer, D. A. Zacharias, S. R. Adams, R. Y. Tsien, *Nat. Biotechnol.* **21**, 1505 (2003).
15. Y. Pommier, *Nat. Rev. Cancer* **6**, 789 (2006).
16. S. Levy et al., *PLoS ONE* **2**, e250 (2007).
17. H. Liu, J. Krizek, A. Bretscher, *Genetics* **132**, 665 (1992).
18. H. Dong, L. Nilsson, C. G. Kurland, *J. Bacteriol.* **177**, 1497 (1995).
19. We thank Z. Yakhini, Z. Kam, R. Milo, S. Sela, and A. Ciechanover for discussions and P. Choukroun for technical assistance. We acknowledge support by the European Research Council, the Israel Science Foundation, and the Kahn Family Foundation. The Weizmann Institute of Science has filed for a patent on the technology described in this report for measuring protein half-lives.

## Supporting Online Material

www.sciencemag.org/cgi/content/full/science.1199784/DC1  
Materials and Methods  
SOM Text  
Figs. S1 to S9  
Tables S1 to S9  
References  
Movie S1

29 October 2010; accepted 16 December 2010

Published online 13 January 2011;

10.1126/science.1199784

# K<sup>+</sup> Channel Mutations in Adrenal Aldosterone-Producing Adenomas and Hereditary Hypertension

Murim Choi,<sup>1</sup> Ute I. Scholl,<sup>1</sup> Peng Yue,<sup>2\*</sup> Peyman Björklund,<sup>3,4\*</sup> Bixiao Zhao,<sup>1\*</sup> Carol Nelson-Williams,<sup>1</sup> Weizhen Ji,<sup>1</sup> Yoonsang Cho,<sup>5</sup> Aniruddh Patel,<sup>1</sup> Clara J. Men,<sup>1</sup> Elias Lolis,<sup>5</sup> Max V. Wisgerhof,<sup>6</sup> David S. Geller,<sup>7</sup> Shrikant Mane,<sup>8</sup> Per Hellman,<sup>4</sup> Gunnar Westin,<sup>4</sup> Göran Åkerström,<sup>4</sup> Wenhui Wang,<sup>2</sup> Tobias Carling,<sup>3</sup> Richard P. Lifton<sup>1†</sup>

Endocrine tumors such as aldosterone-producing adrenal adenomas (APAs), a cause of severe hypertension, feature constitutive hormone production and unrestrained cell proliferation; the mechanisms linking these events are unknown. We identify two recurrent somatic mutations in and near the selectivity filter of the potassium (K<sup>+</sup>) channel KCNJ5 that are present in 8 of 22 human APAs studied. Both produce increased sodium (Na<sup>+</sup>) conductance and cell depolarization, which in adrenal glomerulosa cells produces calcium (Ca<sup>2+</sup>) entry, the signal for aldosterone production and cell proliferation. Similarly, we identify an inherited KCNJ5 mutation that produces increased Na<sup>+</sup> conductance in a Mendelian form of severe aldosteronism and massive bilateral adrenal hyperplasia. These findings explain pathogenesis in a subset of patients with severe hypertension and implicate loss of K<sup>+</sup> channel selectivity in constitutive cell proliferation and hormone production.

**A**ldosterone, a steroid hormone synthesized by the adrenal glomerulosa, is normally produced in two conditions, intravascular volume depletion and hyperkalemia (high plasma K<sup>+</sup> level) (*1*). Volume depletion activates the renin-angiotensin system, producing the hormone angiotensin II (AII), which signals via its

G protein-coupled receptor (GPCR) in glomerulosa cells. The resting membrane potential is set by K<sup>+</sup> channel activity (*2*); both AII signaling and hyperkalemia cause membrane depolarization and activation of voltage-gated Ca<sup>2+</sup> channels. Increased intracellular Ca<sup>2+</sup> provides the normal signal for aldosterone production, and sustained increases lead to glomerulosa cell proliferation (*3–5*); AII also causes increased inositol 1,4,5-trisphosphate (IP<sub>3</sub>) and transient Ca<sup>2+</sup> release from intracellular stores. Aldosterone signaling in the kidney increases electrogenic Na<sup>+</sup> reabsorption, defending intravascular volume, and also increases K<sup>+</sup> secretion.

In primary aldosteronism, the adrenal gland constitutively produces aldosterone in the absence of AII or hyperkalemia, resulting in hypertension and variable hypokalemia (low plasma K<sup>+</sup> level). Primary aldosteronism is found in ~10% of patients referred for evaluation of hypertension. A third or more of these have aldosterone-producing adenoma (APA, also known as Conn's syndrome) of the adrenal cortex (*6*); of the remainder, a small fraction have mutations that cause constitutive

expression of aldosterone synthase (*7*), and the rest are classified as idiopathic.

APAs are typically solitary, well circumscribed, and diagnosed between ages 30 and 70 (*8*). They come to medical attention due to new or worsening hypertension, often with hypokalemia. Aldosterone is elevated while renin levels are suppressed (reflected in a high aldosterone:renin ratio), and a characteristic adrenal mass is seen on computed tomography (CT). Adrenal vein sampling demonstrates predominant aldosterone secretion from the gland harboring the tumor. APAs virtually always remain benign, without local invasion or distant metastasis (*9*). Surgical removal ameliorates or cures hypertension in the large majority of patients (*10*). The mechanisms responsible for neoplasia and cell-autonomous aldosterone production are unknown.

We studied 22 patients with APA (table S1) (*11*). All came to medical attention with hypertension and variable hypokalemia. All had high aldosterone:renin ratios and unilateral adrenal cortical mass on CT. At surgery, adrenocortical tumors of mean diameter 2.8 cm were removed, and pathology in all cases confirmed adrenocortical adenoma.

Genotyping of tumors on Illumina 1M-Duo chips demonstrated two gross classes of tumors: those with zero or few chromosome arms with loss of heterozygosity (LOH) (11 with none, 3 with 1 to 4 LOH events) and those with many large LOH segments (8 with 11 to 19 LOH segments) (table S1 and fig. S1). Subjects with low LOH tumors tended to be younger with smaller tumors.

We performed whole exome capture and Illumina sequencing on four APA-blood pairs from unrelated subjects with no LOH segments. Each tumor sample was assessed by histology to be free of normal adrenal cells; some admixture with blood and stromal cells is inevitable, and we accordingly sequenced samples to high depth of coverage to enable detection of somatic mutations. The mean coverage of each targeted base was 183-fold for blood DNA and 158-fold for tumor DNA, and 97% of all targeted bases in tumor samples were read at least eight times (table S2). We identified high-probability somatic mutations in each tumor ( $P = 10^{-4}$  to  $10^{-56}$  of chance occurrence) (fig. S2), and confirmed each by direct Sanger sequencing

<sup>1</sup>Departments of Genetics and Internal Medicine, Howard Hughes Medical Institute, Yale University School of Medicine, New Haven, CT 06510, USA. <sup>2</sup>Department of Pharmacology, New York Medical College, Valhalla, NY 10595, USA. <sup>3</sup>Department of Surgery, Yale Endocrine Neoplasia Laboratory and Yale Cancer Center, Yale University School of Medicine, New Haven, CT 06510, USA. <sup>4</sup>Department of Surgical Sciences, Uppsala University, Uppsala, Sweden. <sup>5</sup>Department of Pharmacology, Yale University School of Medicine, New Haven, CT 06510, USA. <sup>6</sup>Division of Endocrinology, Henry Ford Hospital, Detroit, MI 48202, USA. <sup>7</sup>Section of Nephrology, Yale University School of Medicine, and Department of Medicine, Veterans Affairs Medical Center, West Haven, CT 06516, USA. <sup>8</sup>Yale Center for Genome Analysis, Yale University School of Medicine, West Haven, CT 06516, USA.

\*These authors contributed equally to this work.

†To whom correspondence should be addressed: E-mail: richard.lifton@yale.edu

**Table 1.** Protein-changing somatic mutations in aldosterone-producing adenomas. Chr, chromosome; Position, position in Human genome build 18; Ref., reference; *P*, significance of difference in frequency of nonreference allele between tumor and blood sequence; *YY1*, transcription factor yin yang 1; *ZNF37*, zinc finger protein 37 homolog; *FZD4*, frizzled 4 homolog; *KCNJ5*,

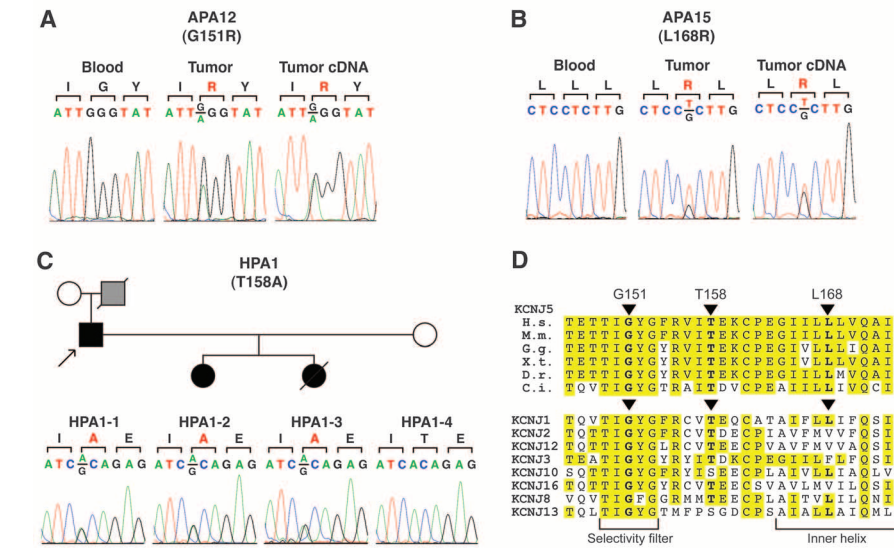
potassium inwardly rectifying channel, subfamily J, member 5; *ARHGAP9*, Rho guanosine triphosphatase activating protein 9; *KDM5C*, lysine (K)-specific demethylase 5C; *PDE9A*, phosphodiesterase 9A; *LRP1B*, low-density lipoprotein receptor-related protein 1B. Mutations in *KCNJ5* in two different tumors are shown in bold font.

Tumor	Chr	Position	Base change	Gene	Effect on protein	No. of reads from tumor			No. of reads from blood		<i>P</i>
						Ref. allele	Non-ref. allele	% of all reads	Ref. allele	Non-ref. allele	
APA9	14	99,813,560	C>G	<i>YY1</i>	T372R	115	69	37.5%	184	0	$1.3 \times 10^{-24}$
	9	114,858,771	C>G	<i>ZFP37</i>	V7L	47	23	32.9%	77	0	$4.0 \times 10^{-9}$
APA12	11	86,341,084	C>A	<i>FZD4</i>	C121F	491	139	22.1%	871	0	$1.6 \times 10^{-55}$
	<b>11</b>	<b>128,286,829</b>	<b>G&gt;A</b>	<b><i>KCNJ5</i></b>	<b>G151R</b>	<b>120</b>	<b>59</b>	<b>33.0%</b>	<b>290</b>	<b>0</b>	<b><math>1.9 \times 10^{-28}</math></b>
APA15	12	56,159,261	G>A	<i>ARHGAP9</i>	R66C	149	65	30.4%	282	1	$1.1 \times 10^{-25}$
	<b>11</b>	<b>128,286,881</b>	<b>T&gt;G</b>	<b><i>KCNJ5</i></b>	<b>L168R</b>	<b>159</b>	<b>65</b>	<b>29.0%</b>	<b>456</b>	<b>0</b>	<b><math>3.5 \times 10^{-35}</math></b>
	X	53,239,430	C>T	<i>KDM5C</i>	V1341M	30	30	50.0%	54	0	$7.6 \times 10^{-11}$
APA22	21	43,054,087	G>A	<i>PDE9A</i>	Exon 13 splice donor GT>AT	90	31	25.6%	123	0	$6.8 \times 10^{-10}$
	2	140,918,376	T>G	<i>LRP1B</i>	R3429S	60	14	18.9%	80	0	$1.7 \times 10^{-5}$

(11). Twelve of 13 putative somatic mutations were confirmed by Sanger sequencing versus 0 of 28 with  $10^{-4} < P < 10^{-3}$  (Table 1, table S3, and fig. S2). The results identified a small number of somatic mutations in each tumor, with a mean of 2.3 protein-altering and 0.8 silent mutations (Table 1 and table S3). Among bases covered  $\geq 40$ -fold (86% of all targeted bases), this represents 0.15 somatic mutations per megabase of exome sequence, a low value compared with a number of malignant tumors (12, 13).

Considering the small number of somatic protein-altering mutations, it was remarkable that one gene, *KCNJ5* (Kir3.4), was mutated in two tumors (Table 1, Fig. 1, and figs. S3 and S4). *KCNJ5* encodes an inwardly rectifying K<sup>+</sup> channel (14). One mutation was G151R, which was present in 33% of tumor reads and none in blood. The other was L168R, present in 29% of tumor reads and none in blood. Each was confirmed as a somatic mutation by Sanger sequencing. Both the wild-type (WT) and mutant *KCNJ5* transcripts were detected in APA cDNA (Fig. 1 and fig. S4). The G151R and L168R mutations are absent in the dbSNP, 1000 Genomes, and Catalogue of Somatic Mutations in Cancer (COSMIC) databases. Sequencing 900 *KCNJ5* alleles from unrelated subjects revealed neither mutation and only two missense variants, R39H and M210I, both in cytoplasmic domains. Staining of normal human adrenal gland with antibodies to *KCNJ5* demonstrated selective staining of zona glomerulosa cells (fig. S5), consistent with tumors arising from mutation in these cells.

Sequencing of *KCNJ5* in the other 18 APA-blood pairs identified six additional somatic mutations. Remarkably, all were either the G151R or L168R mutation. In sum, there were two G151R mutations and six L168R mutations among the



**Fig. 1.** Mutations in *KCNJ5* in aldosterone-producing adenoma and inherited aldosteronism. (A) Sequences of blood and tumor genomic DNA and tumor cDNA of *KCNJ5* codons 150 to 152 in APA12. (B) Sequences of *KCNJ5* codons 167 to 169 in APA15. (C) *KCNJ5* mutation in kindred HPA1. At top, kindred structure is shown; affected members are shown as filled symbols; gray symbol represents a subject who died at age 36 with severe hypertension, suspected to be affected. *KCNJ5* sequences of codons 157 to 159 are shown. Reverse strand traces for (A) to (C) are shown in fig. S4. (D) Conservation of G151, T158, and L168 in orthologs and paralogs. These positions are conserved among chordate orthologs that last shared a common ancestor 750 million years ago. H.s., *Homo sapiens*; M.m., *Mus musculus*; G.g., *Gallus gallus*; X.t., *Xenopus tropicalis*; D.r., *Danio rerio*; C.i., *Ciona intestinalis*. Shown below are the sequences of selected human inward rectifier K<sup>+</sup> channels, demonstrating high conservation among diverse members of this family.

22 tumors (table S1 and fig. S4). The mutations were expressed in tumor cDNA in the six samples studied (fig. S4). Mutant allele frequencies in tumor DNA and cDNA are consistent with mutations being heterozygous in tumor cells. All *KCNJ5* mutations were in the low LOH

group, including 7 of the 11 tumors with no LOH segments.

Even using an inflated estimate of one somatic mutation per million base pairs in these tumors, the probability of seeing either of two somatic mutations recur by chance in 6 of 20 other

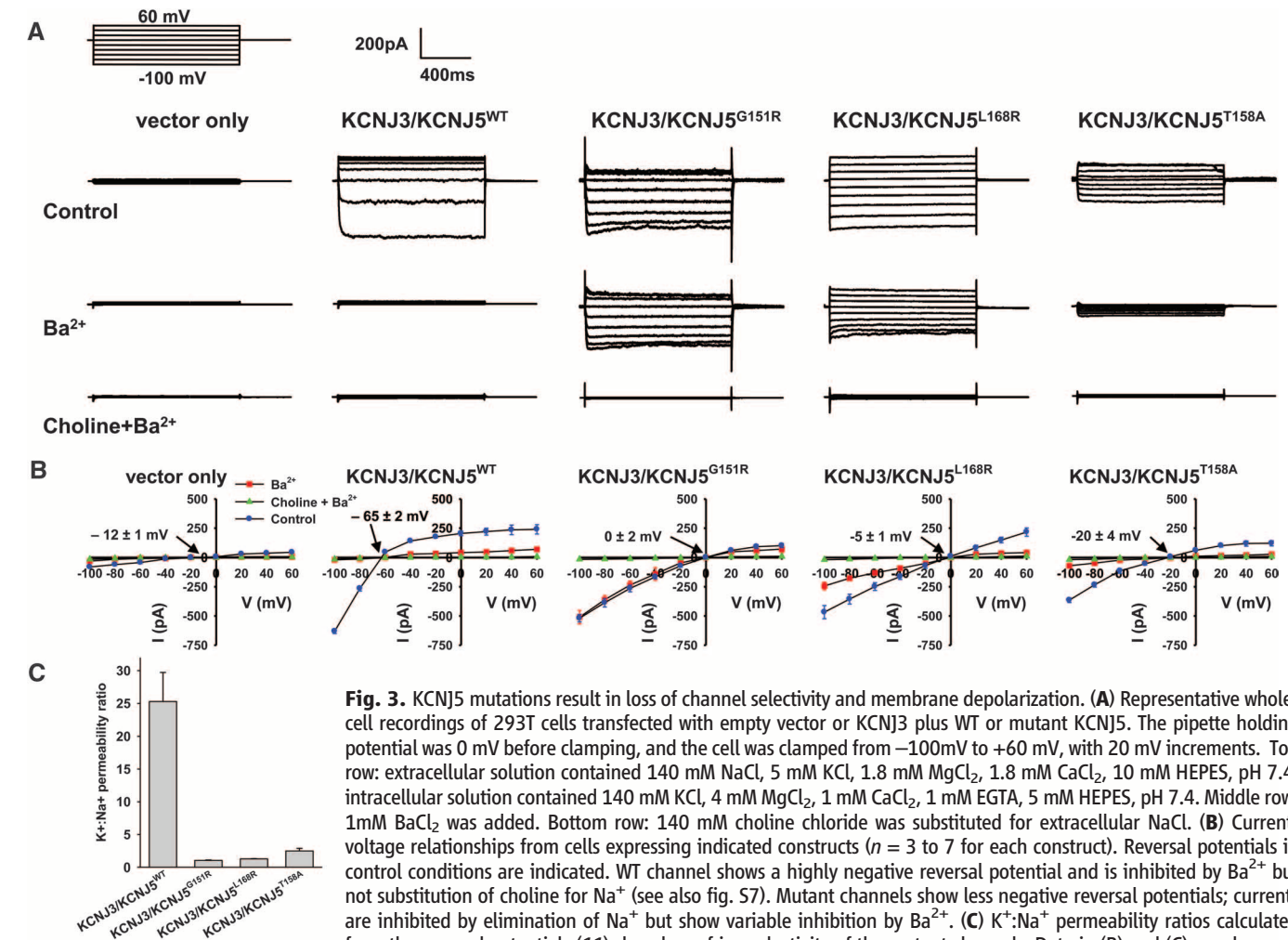
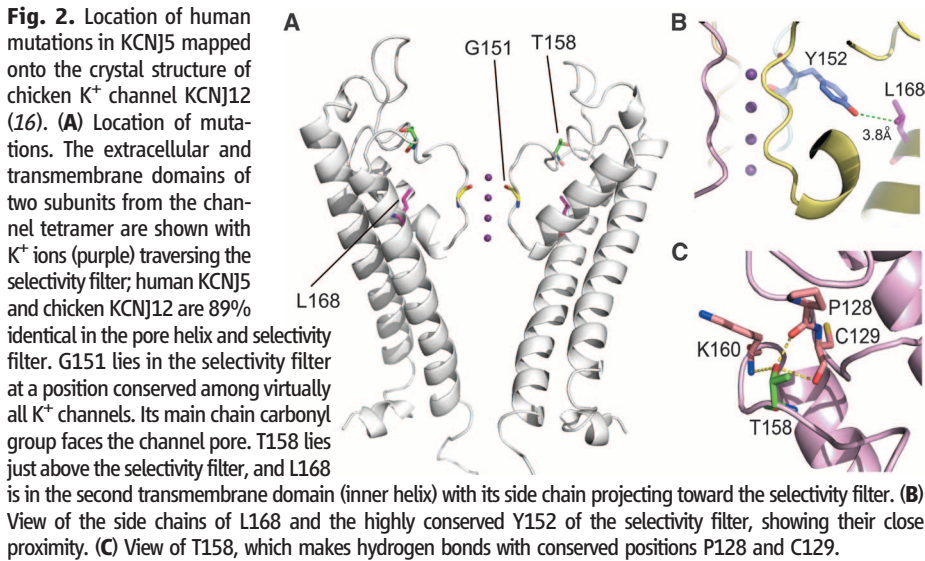


tumors is  $<10^{-30}$ , strongly implicating these two mutations in the pathogenesis of APA. The recurrence of the identical mutations strongly implies a genetic gain-of-function mechanism.

The crystal structures of a number of  $K^+$  channels have been determined, and the general features are highly conserved (15); the closest to KCNJ5 is chicken KCNJ12, another inward

rectifier (16). The wild-type amino acids, G151 and L168, lie at highly conserved positions, which we mapped onto the KCNJ12 structure. G151 is the first glycine of the GYG motif of the  $K^+$  channel selectivity filter (Figs. 1D and 2A); glycine at this position is found in virtually every  $K^+$  channel in the biological world (17, 18). The main chain carbonyl groups of G151 face the pore in the channel tetramer (Fig. 2A), and their distances from one another approximate the distances of oxygen atoms in the hydration shell surrounding  $K^+$  ions, stripping water from the ion (15, 16). L168 is also conserved among KCNJ5 orthologs and inward rectifiers (Fig. 1D). L168 lies in the second transmembrane domain (inner helix) of KCNJ5; its side chain abuts the highly conserved tyrosine side chain of the GYG motif (Fig. 2B).

Previous studies have shown that mutations in and near  $K^+$  channel selectivity filters can alter channel selectivity to produce nonselective cation channels (17, 19). KCNJ5 exists both as homotetramers and heterotetramers with KCNJ3 (KCNJ3 is inactive as a homotetramer) (20); heterotetramers are more active than homotet-



**Fig. 3.** KCNJ5 mutations result in loss of channel selectivity and membrane depolarization. (A) Representative whole-cell recordings of 293T cells transfected with empty vector or KCNJ3 plus WT or mutant KCNJ5. The pipette holding potential was 0 mV before clamping, and the cell was clamped from  $-100$  mV to  $+60$  mV, with  $20$  mV increments. Top row: extracellular solution contained  $140$  mM NaCl,  $5$  mM KCl,  $1.8$  mM MgCl<sub>2</sub>,  $1.8$  mM CaCl<sub>2</sub>,  $10$  mM HEPES, pH  $7.4$ ; intracellular solution contained  $140$  mM KCl,  $4$  mM MgCl<sub>2</sub>,  $1$  mM CaCl<sub>2</sub>,  $1$  mM EGTA,  $5$  mM HEPES, pH  $7.4$ . Middle row:  $1$  mM BaCl<sub>2</sub> was added. Bottom row:  $140$  mM choline chloride was substituted for extracellular NaCl. (B) Current-voltage relationships from cells expressing indicated constructs ( $n = 3$  to  $7$  for each construct). Reversal potentials in control conditions are indicated. WT channel shows a highly negative reversal potential and is inhibited by Ba<sup>2+</sup> but not substitution of choline for Na<sup>+</sup> (see also fig. S7). Mutant channels show less negative reversal potentials; currents are inhibited by elimination of Na<sup>+</sup> but show variable inhibition by Ba<sup>2+</sup>. (C) K<sup>+</sup>:Na<sup>+</sup> permeability ratios calculated from the reversal potentials (11) show loss of ion selectivity of the mutant channels. Data in (B) and (C) are shown as mean  $\pm$  SEM. Reversal potentials and K<sup>+</sup>:Na<sup>+</sup> permeability ratios are significantly different between wild-type and mutant channels ( $P < 0.01$  by Student's  $t$  test).

ramers, and activity can be increased by activation of GPCRs such as dopamine D2 (14, 21). Both *KCNJ5* and *KCNJ3* are expressed in human adrenal cortex (table S4). We expressed wild-type or mutant *KCNJ5* with *KCNJ3* in 293T cells (Fig. 3 and fig. S6) and measured currents at voltages from  $-100$  mV to  $+60$  mV using perforated whole-cell recording with physiologic solutions including  $140$  mM  $K^+$  inside and  $5$  mM  $K^+$ ,  $140$  mM  $Na^+$  outside the cell (11). *KCNJ3/KCNJ5*<sup>WT</sup> induced a robust inwardly rectifying  $Ba^{2+}$ -sensitive current that hyperpolarized the membrane with a reversal potential of  $-65 \pm 2$  mV (Fig. 3). Co-expression with the dopamine D2 receptor and addition of dopamine increased current by  $\sim 50\%$  (fig. S7, A and B). These are all characteristic features of *KCNJ3/KCNJ5* heterotetramers (14). The  $K^+ : Na^+$  permeability ratio, estimated from the Goldman equation, was  $25.3 \pm 4.4:1$ . In contrast, *KCNJ3/KCNJ5*<sup>G151R</sup> channels produced currents that showed loss of inhibition by  $Ba^{2+}$  and membrane depolarization with a shift of the reversal potential to  $0 \pm 2$  mV. This depolarization is attributable to increased  $Na^+$  conductance: whereas substitution of choline for  $Na^+$  had no effect on the WT channel, elimination of  $Na^+$  markedly inhibited *KCNJ3/KCNJ5*<sup>G151R</sup> currents either with (Fig. 3) or without (fig. S7, C and D)  $Ba^{2+}$ . The calculated  $K^+ : Na^+$  permeability ratio is diminished to  $1.0 \pm 0.1:1$ , consistent with loss of channel selectivity. This loss of ion selectivity is similar to effects seen with other selectivity filter mutations (17). *KCNJ3/KCNJ5*<sup>L168R</sup> channels behave similarly, producing a reversal potential of  $-5 \pm 1$  mV and a  $K^+ : Na^+$  permeability ratio of  $1.3 \pm 0.1:1$  (Fig. 3). Similar results were obtained with *KCNJ5* homotetramers (fig. S8). In glomerulosa cells, membrane depolarization activates voltage-gated  $Ca^{2+}$  channels, increasing intracellular  $Ca^{2+}$ , thereby increasing aldosterone production (Fig. 4) (1). Similarly, chronic  $Ca^{2+}$  stimulation promotes increased proliferation in glomerulosa (3–5) and other cell types (22, 23), which can account for clonal expansion of cells

harboring these somatic mutations and adenoma formation.

These inferences from somatic mutations in tumors suggest that inherited mutations in *KCNJ5* with similar effect could cause a Mendelian form of primary aldosteronism with bilateral adrenal hyperplasia, because in this case every adrenal cell would harbor the mutation. We recently described just such a syndrome of unknown cause in a father and his two daughters who were all diagnosed between ages 4 and 7 with severe hypertension, aldosteronism, and massive adrenal hyperplasia (table S5) (24). All three individuals had a radical intervention, bilateral adrenalectomy in childhood. Pathology demonstrated massive hyperplasia of the adrenal cortex (paired adrenal weights up to  $81$  g; normal  $<12$  g). The sequence of *KCNJ5* identified a heterozygous T158A mutation that cosegregated with the disease (Fig. 1C and fig. S4I). This variant is absent in the dbSNP and 1000 Genomes databases and in 900 control alleles. This threonine is conserved among *KCNJ5* orthologs and other inward rectifiers (Fig. 1D) and lies in the loop between the selectivity filter and the second transmembrane domain; its hydroxyl group hydrogen bonds with conserved residues in the loop between the first transmembrane domain and the pore helix, constraining the structure (Fig. 2C). The T158A mutation eliminates these hydrogen bonds. Similar to the other mutations, *KCNJ3/KCNJ5*<sup>T158A</sup> channels showed reduced selectivity ( $K^+ : Na^+$  permeability ratio of  $2.5 \pm 0.4:1$ ) and membrane depolarization, with a reversal potential of  $-20 \pm 4$  mV (Fig. 3B). Similar results were seen in homotetramers (fig. S8).

These findings implicate inherited and acquired mutations in *KCNJ5* in aldosteronism associated with cell autonomous proliferation. The very small number of somatic mutations observed, the young age of many APA subjects with *KCNJ5* mutations (four of eight under age 35), and Mendelian transmission of the inherited syndrome are consistent with the *KCNJ5* mutations being sufficient for both constitutive aldosterone secretion

and cell proliferation. The increased  $Na^+$  conductance and membrane depolarization resulting from these mutations implicate activation of voltage-gated  $Ca^{2+}$  channels in the pathophysiologic mechanism (Fig. 4) (1, 3–5).

The effects of these mutations in and near the selectivity filter to reduce channel selectivity are consistent with previous *in vitro* studies (17, 19). In addition, mutation of the homologous glycine to serine in *KCNJ6* in the *weaver* mouse also produces a  $Na^+$ -conducting channel, leading to selective loss of neurons in cerebellum and substantia nigra (25). Glomerulosa cells have constitutively open “leak”  $K^+$  channels and a high  $Na^+ / K^+$  adenosine triphosphatase activity (26); such differences may contribute to different fates in these cell types.

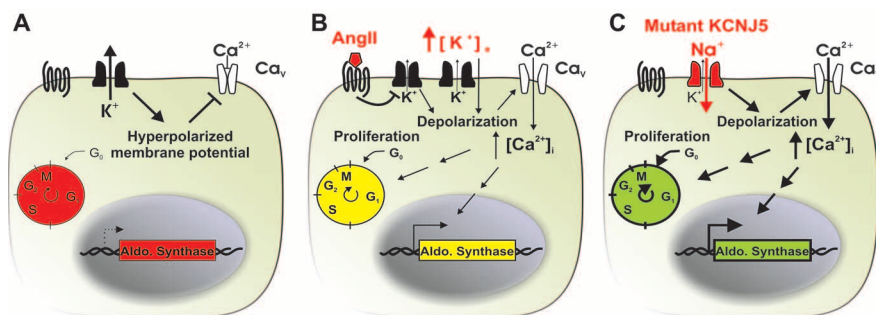
Because mutations in and near  $K^+$  channel selectivity filters can alter ion selectivity (19), the restricted spectrum of mutations found in APAs is noteworthy. One possible explanation is that mutant channels must cause sufficient  $Na^+$  permeability for tumor development but not so great as to cause cell death (23, 25); these requirements may restrict the mutational spectrum. The lower relative  $Na^+$  permeability observed with the inherited T158A mutation is consistent with allelic variation in effect. It will be of interest to determine the prevalence and spectrum of *KCNJ5* mutations in other cohorts of patients with APAs and with unexplained aldosteronism.

These findings also raise the question of the normal role of *KCNJ5* in glomerulosa cells. In rodent and cow, members of the “leak”  $K^+$  channel family (*KCNK2*, *KCNK3*, and *KCNK9*) appear to set the resting potential (27, 28). Dopamine, an inhibitor of aldosterone release, increases activity of  $K^+$  channels containing *KCNJ5* (21), suggesting that *KCNJ5* may normally inhibit aldosterone production. We sequenced *KCNK2*, *KCNK3*, and *KCNK9* in the tumor cohort and found no mutations, consistent with *KCNJ5* having a privileged role in producing APA.

Lastly, these findings demonstrate a role for ion channel mutations in neoplasia. The distinct mechanism of these *KCNJ5* mutations may be related to the benign nature of these tumors. It will be of interest to determine whether other endocrine neoplasias have related mutations that account for concomitant cell proliferation and hormone release. Mutations in other  $K^+$  channel genes have been identified in various human cancers, but their importance has been uncertain. These include mutations altering conserved residues in or near voltage-regulating segments in *KCNB2*, *KCNC2*, and *KCNQ5* from glioblastoma, breast cancer, and colorectal cancers (12, 13, 29). Investigation of the functional consequences of these mutations will be of interest.

#### References and Notes

1. A. Spät, L. Hunyady, *Physiol. Rev.* **84**, 489 (2004).
2. A. Spät, *Mol. Cell. Endocrinol.* **217**, 23 (2004).
3. P. E. McEwan, G. B. Lindop, C. J. Kenyon, *Am. J. Physiol.* **271**, E192 (1996).



**Fig. 4.** Proposed mechanism underlying aldosterone-producing adenoma and Mendelian aldosteronism. (A) Adrenal glomerulosa cells have a high resting  $K^+$  conductance, which produces a highly negative membrane potential (2). (B) Membrane depolarization by either elevation of extracellular  $K^+$  or closure of  $K^+$  channels by angiotensin II activates voltage-gated  $Ca^{2+}$  channels, increasing intracellular  $Ca^{2+}$  levels (1). This provides signals for increased expression of enzymes required for aldosterone biosynthesis, such as aldosterone synthase, and for increased cell proliferation. (C) Channels containing *KCNJ5* with G151R, T158A, or L168R mutations conduct  $Na^+$ , resulting in  $Na^+$  entry, chronic depolarization, constitutive aldosterone production, and cell proliferation.

4. M. Pawlikowski, A. Gruszka, S. Mucha, G. Melen-Mucha, *Endocr. Regul.* **35**, 139 (2001).
5. A. Tanabe *et al.*, *J. Endocrinol. Invest.* **21**, 668 (1998).
6. G. P. Rossi *et al.*; PAPY Study Investigators, *J. Am. Coll. Cardiol.* **48**, 2293 (2006).
7. R. P. Lifton *et al.*, *Nature* **355**, 262 (1992).
8. V. Kumar, A. K. Abbas, N. Fausto, J. C. Aster, Eds., in *Robbins and Cotran Pathologic Basis of Disease* (Saunders, Philadelphia, ed. 8, 2009), chap. 24.
9. R. P. Ghose, P. M. Hall, E. L. Bravo, *Ann. Intern. Med.* **131**, 105 (1999).
10. J. M. Calvo-Romero, J. L. Ramos-Salado, *Postgrad. Med. J.* **76**, 160 (2000).
11. Materials and methods are available as supporting material on Science Online.
12. L. D. Wood *et al.*, *Science* **318**, 1108 (2007).
13. T. Sjöblom *et al.*, *Science* **314**, 268 (2006).
14. G. Krapivinsky *et al.*, *Nature* **374**, 135 (1995).
15. D. A. Doyle *et al.*, *Science* **280**, 69 (1998).
16. X. Tao, J. L. Avalos, J. Chen, R. MacKinnon, *Science* **326**, 1668 (2009).
17. L. Heginbotham, Z. Lu, T. Abramson, R. MacKinnon, *Biophys. J.* **66**, 1061 (1994).
18. B. Roux, *Annu. Rev. Biophys. Biomol. Struct.* **34**, 153 (2005).
19. K. M. Dibb *et al.*, *J. Biol. Chem.* **278**, 49537 (2003).
20. S. Corey, D. E. Clapham, *J. Biol. Chem.* **273**, 27499 (1998).
21. K. A. Gregerson *et al.*, *Endocrinology* **142**, 2820 (2001).
22. C. R. Kahl, A. R. Means, *Endocr. Rev.* **24**, 719 (2003).
23. H. L. Roderick, S. J. Cook, *Nat. Rev. Cancer* **8**, 361 (2008).
24. D. S. Geller *et al.*, *J. Clin. Endocrinol. Metab.* **93**, 3117 (2008).
25. B. Navarro *et al.*, *Science* **272**, 1950 (1996).
26. G. Hajnóczky *et al.*, *Endocrinology* **130**, 1637 (1992).
27. J. J. Enyeart, L. Xu, S. Danthi, J. A. Enyeart, *J. Biol. Chem.* **277**, 49186 (2002).
28. G. Cziráj, P. Enyedi, *Mol. Endocrinol.* **16**, 621 (2002).
29. D. W. Parsons *et al.*, *Science* **321**, 1807 (2008).
30. We thank the patients whose participation made this study possible and the staff of the Yale West Campus Genomics Center and the Endocrine Surgical Laboratory, Clinical Research Centre, University Hospital, Uppsala. Supported in

part by the Fondation Leducq Transatlantic Network in Hypertension, National Institutes of Health (NIH) grant DK54983, the Yale Center for Human Genetics and Genomics, Yale NIH O'Brien Center for Kidney Research, and the Yale NIH Clinical Translational Science Award, and by the Swedish Cancer Society, the Swedish Research Council, and the Lions Cancer Fund, Uppsala. U.I.S. is a fellow of the Deutsche Forschungsgemeinschaft; B.Z. is an investigator of the Yale Medical Scientist Training Program; T.C. is a Doris Duke–Damon Runyon Clinical Investigator; R.P.L. is a paid scientific advisor to Merck and is an investigator of the Howard Hughes Medical Institute.

#### Supporting Online Material

[www.sciencemag.org/cgi/content/full/331/6018/768/DC1](http://www.sciencemag.org/cgi/content/full/331/6018/768/DC1)

Materials and Methods

Figs. S1 to S8

Tables S1 to S5

References

7 October 2010; accepted 3 January 2011

10.1126/science.1198785

# Retrieval Practice Produces More Learning than Elaborative Studying with Concept Mapping

Jeffrey D. Karpicke\* and Janell R. Blunt

Educators rely heavily on learning activities that encourage elaborative studying, whereas activities that require students to practice retrieving and reconstructing knowledge are used less frequently. Here, we show that practicing retrieval produces greater gains in meaningful learning than elaborative studying with concept mapping. The advantage of retrieval practice generalized across texts identical to those commonly found in science education. The advantage of retrieval practice was observed with test questions that assessed comprehension and required students to make inferences. The advantage of retrieval practice occurred even when the criterial test involved creating concept maps. Our findings support the theory that retrieval practice enhances learning by retrieval-specific mechanisms rather than by elaborative study processes. Retrieval practice is an effective tool to promote conceptual learning about science.

Most thought on human learning is guided by a few tacit assumptions. One assumption is that learning happens primarily when people encode knowledge and experiences. A related assumption is that retrieval—the active, cue-driven process of reconstructing knowledge—only measures the products of a previous learning experience but does not itself produce learning. Just as we assume that the act of measuring a physical object would not change the size, shape, or weight of the object, so too people often assume that the act of measuring memory does not change memory (1, 2). Thus, most educational research and practice has focused on enhancing the processing that occurs when students encode knowledge—that is, getting knowledge “in memory.” Far less attention has been paid to the potential importance of retrieval to the process of learning. Indeed, recent National Research Council books

about how students learn in educational settings (3–5) contain no mention of retrieval processes.

It is beyond question that activities that promote effective encoding, known as elaborative study tasks, are important for learning (6). However, research in cognitive science has challenged the assumption that retrieval is neutral and uninfluential in the learning process (7–11). Not only does retrieval produce learning, but a retrieval event may actually represent a more powerful learning activity than an encoding event. This research suggests a conceptualization of mind and learning that is different from one in which encoding places knowledge in memory and retrieval simply accesses that stored knowledge. Because each act of retrieval changes memory, the act of reconstructing knowledge must be considered essential to the process of learning.

Most previous research on retrieval practice has been conducted in the verbal learning tradition of memory research (12). The materials used have often not reflected the complex information students learn in actual educational settings (13). Most previous research has not

used assessments thought to measure meaningful learning, which refers to students' abilities to make inferences and exhibit deep understanding of concepts (14, 15). Perhaps the greatest impediment to broad application of retrieval practice, though, is that we do not know whether retrieval activities are more effective than other active, elaborative learning activities. Retrieval practice might produce levels of learning that are essentially the same as those produced by elaborative studying. Alternatively, if there are retrieval-specific mechanisms that promote learning, then retrieval practice may represent a way to promote student learning that goes beyond elaborative study activities used in science education.

The present experiments put retrieval practice to a test. Elaborative learning activities hold a central place in contemporary education. We examined the effectiveness of retrieval practice relative to elaborative studying with concept mapping (16–18). In concept mapping, students construct a diagram in which nodes are used to represent concepts, and links connecting the nodes represent relations among the concepts. Concept mapping is considered an active learning task, and it serves as an elaborative study activity when students construct concept maps in the presence of the materials they are learning. Under these conditions, concept mapping bears the defining characteristics of an elaborative study method: It requires students to enrich the material they are studying and encode meaningful relationships among concepts within an organized knowledge structure.

In two experiments, we compared the effectiveness of retrieval practice and elaborative studying with concept mapping for producing meaningful learning of science materials. Eighty undergraduate students participated in Experiment 1. The students first studied a science text under one of four conditions within a single initial learning session. In the study-once condition, students studied the text in a single study period. In the repeated study condition, students studied the text in four consecutive study periods (8). In the elaborative concept mapping condition, students studied the

Department of Psychological Sciences, Purdue University, West Lafayette, IN 47907, USA.

\*To whom correspondence should be addressed. E-mail: [karpicke@purdue.edu](mailto:karpicke@purdue.edu)